Benjamin Tjoa *et al.* Appl. No. 10/789,807 Response dated October 24, 2005 Reply to Office Action of August 23, 2005

## AMENDMENTS TO THE SPECIFICATION:

Please replace paragraph 46 with the following amended paragraph:

Suitable predetermined antigens for use in the present invention can include any antigen for which T-cell activation is desired. Such antigens can include, for example, bacterial cells, or other preparation comprising bacterial antigens, tumor specific or tumor associated antigens (e.g., whole tumor or cancer cells, a tumor cell lysate, tumor cell membrane preparations, isolated or partially isolated antigens from tumors, fusion proteins, liposomes, and the like), viral particles or other preparations comprising viral antigens, and any other antigen or fragment of an antigen, e.g., a peptide or polypeptide antigen. In certain embodiments, the antigen can be associated with prostate cancer, for example the antigen can be, but not limited to, prostate specific membrane antigen (PSMA), prostatic acid phosphatase (PAP), or prostate specific antigen (PSA). (See, e.g., Pepsidero et al., Cancer Res. 40:2428-32 (1980); McCormack et al., Urology 45:729-44 (1995).) The antigen can also be a bacterial cell, bacterial lysate, membrane fragment from a cellular lysate, or any other source known in the art. The antigen can be expressed or produced recombinantly, or even chemically synthesized. The recombinant antigen can also be expressed on the surface of a host cell (e.g., bacteria, yeast, insect, vertebrate or mammalian cells), can be present in a lysate, or can be purified from the lysate. Alternatively, the antigen can be encoded by nucleic acids which can be ribonucleoic ribonucleic acid (RNA) or deoxyribonucleic acid (DNA), that are purified or amplified from a tumor cell.

Please replace paragraph 82 with the following amended paragraph:

Each of the populations of monocytes, activated and non-activatedwere non-activated were then incubated in X-VIVO-15<sup>®</sup> with 2% HSA in the presence of GM-CSF alone or in combination with IL-4 for 5 days. The resulting immature DCs were loaded with influenza A M1-A4 40mer peptide or keyhole limpet hemocyanin (KLH) for one hour prior to washing and maturing with BCG (1:400 dil) and IFN-γ (500 U/mL). After harvesting and washing the

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mature DC, co-cultures with DCs and autologous PBMCs were set up at a 1:10 DC:PBMC ratio in AIM-V<sup>®</sup> plus 5% human AB sera (HuAB Sera) supplemented with 20 ng/ml IL-2 from day 2 through day 8. After eight days of culture the T cell lines were harvested and analyzed for M1-A4 specific CD8 T cell expansion (V $\beta$ 17<sup>+</sup> CD8<sup>+</sup> T cells) by flow cytometry.